

Routes of influenza transmission

Ben Killingley, Jonathan Nguyen-Van-Tam

University of Nottingham, Nottingham, UK.

Correspondence: Ben Killingley, Division of Epidemiology and Public Health, Clinical Sciences Building, City Hospital Campus, Hucknall Road, Nottingham NG5 1PB, UK.

Email: Ben.Killingley@nottingham.ac.uk

Remarkably little is known definitively about the modes of influenza transmission. Thus, important health policy and infection control issues remain unresolved. These shortcomings have been exposed in national and international pandemic preparedness activities over recent years. Indeed, WHO, CDC, ECDC and the U.S. Institute of Medicine have prioritised understanding the modes of influenza transmission as a critical need for pandemic planning. Studying influenza transmission is difficult; seasonality, unpredictable attack rates, role of environmental parameters such as temperature and humidity, numbers of participants required and confounding variables all present considerable obstacles to the execution of definitive studies. A range of investigations performed to date have failed to

provide definitive answers and key questions remain. Reasons for this include the fact that many studies have not sought to investigate routes of transmission as a primary objective (instead, they have evaluated specific interventions) and that fieldwork in natural settings, specifically assessing the dynamics and determinants of transmission between humans, has been limited. The available evidence suggests that all routes of transmission (droplet, aerosol and contact) have a role to play; their relative significance will depend on the set of circumstances acting at a given time. Dictating the process are factors related to the virus itself, the host and the environment.

Keywords Aerosol, contact, droplet, influenza, transmission.

Please cite this paper as: Killingley and Nguyen-Van-Tam (2013) Routes of influenza transmission. *Influenza and Other Respiratory Viruses* 7(Suppl. 2), 42–51.

Introduction

Limited understanding of influenza transmission has been a frequent obstacle during the development of pandemic influenza infection prevention and mitigation strategies. The science is hotly debated, especially the relative importance of transmission via large droplets and droplet nuclei.^{1,2} In the aftermath of the 2009 A (H1N1) pandemic, clarification of the relative importance of different modes of transmission is critical for the refinement of evidence-based infection control advice for healthcare settings, schools, workplaces and homes.

Transmission of an infectious disease is the process by which an infectious organism moves from one host to another and causes disease. There are many factors that contribute to and influence this process and to appreciate them one must first understand the basic pathophysiology of the underlying disease process.

Influenza replicates in epithelial cells throughout the respiratory tree (both upper and lower respiratory tract).³ Human viruses preferentially bind to cell surface receptors (sialyloligosaccharides) terminated by a N-acetylsialic acid linked to galactose by an $\alpha(2,6)$ -linkage.⁴ The predominance of these receptors in different tissues reflects the tropism

seen, for example $\alpha(2,6)$ are found mainly in the human respiratory tract.⁵ As a result, both virus entry and exit in humans occurs through the respiratory tract, that is, mouth and nose. Virus is released from an infected host during events such as coughing, sneezing and talking. An 'expiratory spray' of different sized particles in which virus travels is produced. Virus gains entry to a new host via inhalation and/or direct contact and/or indirect contact. From here, target epithelial cells can be reached. The potential of the conjunctivae to mediate transmission of human influenza viruses remains uncertain⁶ although data from tropism experiments with pandemic H1N1⁷ and outbreaks of avian H7 viruses in humans that are marked by conjunctivitis⁸ confirm the presence of $\alpha(2,3)$ receptors in the eye. Furthermore, it has recently been shown that an aerosolised live attenuated virus can reach the nasopharynx via an ocular route.⁹ There is very little evidence to suggest that the faecal-oral or waterborne route of transmission occurs in humans, in contrast to transmission that occurs amongst birds.^{10,11}

Three routes of human influenza infection transmission are widely accepted:

- Droplets: these particles can deposit on mucous surfaces of the upper respiratory tract (URT) such as the mouth

and nose. They can be inhaled but are too large ($>10\ \mu\text{m}$) to reach the lungs.

- Droplet nuclei (hereafter referred to as aerosols): these particles are small enough to be inhaled ($<5\ \mu\text{m}$) and reach the lower respiratory tract (LRT). They may also deposit on surfaces in the URT.
- Contact transmission: particles are transferred to mucous membranes of the upper respiratory tract either directly or via a contaminated object or person, that is, indirectly.

For viruses to cause infection in new hosts, a number of prerequisites exist; (i) they must survive in the environment; (ii) they must reach target cells in a new host; and (iii) enough virus must reach target cells such that an infectious dose is achieved and infection initiated.

By considering the transmission pathways outlined above, it is evident that factors related to the virus, the environment and the host may all contribute to transmission (Figure 1). To formulate and implement effective influenza control measures such as personal hygiene, social distancing and infection control, it is critical to understand the above factors as each of these in turn can influence the route(s) of transmission that are active.

Nearly, a century has passed since the first studies of influenza transmission were conducted and many questions remain unanswered, for example;

- What is the relative significance of the different routes of influenza transmission?
- Do transmission routes differ in different settings?

- What is the extent and significance of virus deposition in the environment?
- What environmental factors influence transmission?
- What is the relative effectiveness of hand hygiene, surgical face masks (SFM) and respirators in preventing transmission?
- What other interventions may be used to reduce transmission?
- How important is transmission from asymptomatic and pre-symptomatic individuals?

This article is a review of the biological and scientific determinants that contribute to an understanding of the routes of transmission that operate in humans. For each route, evidence for and against its significance is presented. It concludes by identifying ongoing research gaps.

Evidence base

The evidence base on influenza transmission is largely derived from six categories of study. Each of these has been evaluated to see whether or not a significant role for each route of transmission is supported.

1. Studies assessing influenza virus deposition and survival in the environment that inform the biologic plausibility of the proposed routes of transmission
2. Studies examining the epidemiology of disease in closed or semi-closed settings
3. Non-pharmaceutical intervention studies
4. Human influenza challenge studies

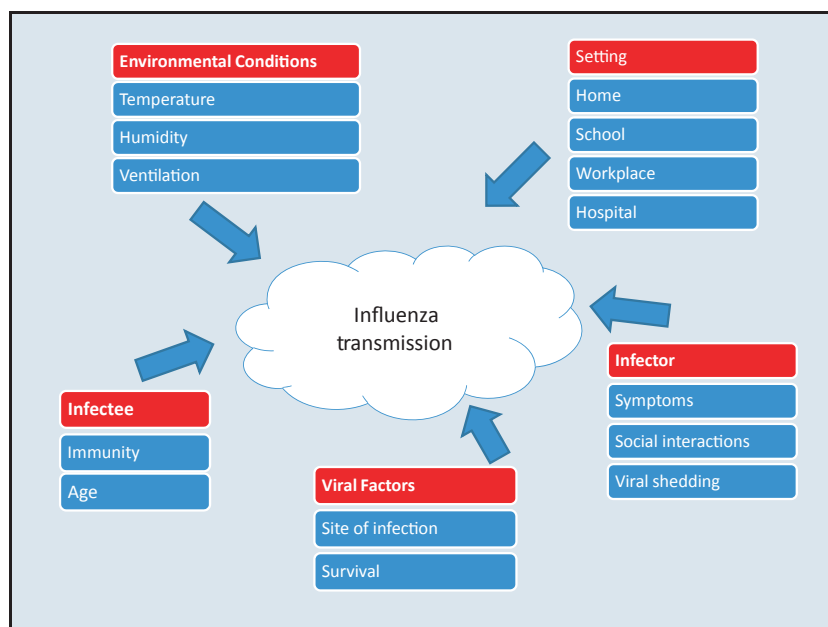


Figure 1. Factors that affect influenza transmission.

5. Animal models of transmission
6. Transmission modelling

Studies assessing influenza virus deposition and survival in the environment that inform the biologic plausibility of the proposed routes of transmission

Contact transmission

There is sound evidence supporting influenza virus survival on fomites^{12–14} and hands^{12,15,16} for periods consistent with the possibility of onward transmission. Data regarding the likely survival time of virus deposited on surfaces are relatively heterogeneous and factors such as virus concentration of the inoculum, type of surface and temperature and humidity all affect virus survival. Thus, it is not possible to provide absolute numbers or ranges for survival times further than to say that estimates lie in the range of a few hours to several days. In general, the data support longer survival on hard (non-porous) surfaces than on softer (porous) items.

Few data demonstrate the recovery of viable virus from hands or surfaces contaminated by patients with influenza in natural settings.^{17,18} Detecting virus by PCR is more sensitive, but whilst swab positive rates in some studies have been high (20–50%)^{19,20}, others have found lower rates 2–5%.^{17,21,22} This might suggest that virus deposited by infected patients does not contaminate the vast majority of fomites in high titre. However, it might also reflect limitations in sampling efficiency, study designs and/or virological techniques.

Consideration of the transmission pathway for the indirect contact route does raise doubt about its plausibility. How likely is it that an infectious dose of virus can persist whilst passing along the transmission chain, infected secretions → (fingertips of infectors →) fomites → fingertips of infectees → mucous membranes → initiation of infection? No direct evidence exists to show that the contact route can mediate transmission, and the data currently available do not fully support the contact route of transmission playing a significant role in the spread of influenza.

Droplet transmission

Coughing and sneezing produce 'expiratory sprays' that consist of a range of particles lying on a size continuum from large particles (droplets) to small particles (aerosols) (reviewed by Nicas²³). Aerobiological studies reveal that the vast majority of pathogens excreted in expiratory sprays are contained within droplets; this is related to the fact that droplets constitute 99% of the volume of an expiratory spray.^{23–26} These particles behave ballistically and fall out of circulation within a few feet (range is proportional to size); they are potentially inhalable but cannot reach the LRT. Initiation of infection following the inhalation of particles is

dependent on several factors such as infectious dose [thought to be higher in the URT than the LRT²], nose or mouth breathing, tidal volume, breathing rate and timing – so that an inspiratory breath in a susceptible contact occurs immediately after particle generation by an infected case. So, whilst the basic concept of droplet transmission may at first be readily accepted, the constraining factors mentioned have led some to consider it a rare event.²⁷

Aerosol transmission

The majority of particles produced by an infected individual are <5 µm. Somewhat paradoxically, only a minor proportion of the total pathogens excreted will be contained within such particles, perhaps as low as 1%; this is a reflection of their relative volume.²⁵ By inference, the likelihood of infectious aerosol particles being produced is probably increased in patients who are shedding higher virus titres (e.g. those in the early days of illness, children, immunocompromised patients^{28–31}), and data in support of this are emerging (Werner Bischoff 2012, Personal Communication).

Detecting the presence of influenza in the air is the first step in a chain of evidence needed to confirm that influenza viruses, emitted from an infected individual and existing as infectious aerosols, can initiate infection in a person exposed to them. The other steps in this sequence are (i) confirming that live virus is present and (ii) confirming that inhaled live virus can initiate infection.

Evidence backing up at least the potential for bioaerosol transmission of influenza is accumulating. Supporting evidence comes from the detection of influenza virus (by PCR) in the air of natural settings.^{17,32–35} More recent work has shown that viable virus can be detected in exhaled breath³⁶ and cough samples from infected individuals³⁷ and that significant heterogeneity exists between individuals in the amount of virus emitted.³⁷ Influenza can survive in air for periods long enough to allow transmission (reviewed by Weber⁶). Overall investigators find that survival is prolonged (up to 24 hours) at low relative humidity.

Studies examining the epidemiology of disease in closed or semi-closed settings

Most outbreak studies are inconclusive in determining the relative importance of different modes of influenza transmission. They suggest that most influenza transmission occurs at close range.^{38–50} Although there is little epidemiological data to support long-range transmission of influenza, these data need to be placed in the context of the rapid diminution of concentrations of infectious aerosols as distance from the generating source increases. Thus, the absence of evidence for long-range transmission does not preclude a significant role for short-range spread via aerosol-sized particles, in some circumstances, at ranges normally or traditionally attributed to only ballistic-sized larger droplets.

Two reports describe circumstances favourable to aerosol transmission and its likely occurrence. One occurred aboard an aircraft that was grounded for 4.5 hours and had the ventilation system shut down⁴⁷, and the other took place on a hospital ward where the flow of air had been inadvertently altered.⁴⁸

Non-pharmaceutical intervention studies

A role for contact spread in the transmission of respiratory infections is supported by three systematic reviews^{51–53} and one meta-analysis⁵⁴ that included data on hand hygiene (HH) to reduce the spread of acute respiratory infections. One review was specific to influenza⁵¹, but in general these papers relate to acute respiratory infections as a whole as there is little organism-specific data. All reviews comment on the heterogeneity and often poor quality of studies performed, but all conclude that HH can reduce episodes of respiratory illness. More recently two studies have shown that HH interventions can reduce the incidence of influenza-like illness (ILI) and laboratory-confirmed influenza.^{55,56}

Surgical face masks (SFM) present a barrier to droplet transmission and by virtue of covering the mouth and nose can also reduce hand-to-face contact transmission. Respirators have the added potential benefit of reducing aerosol transmission as they can filter out droplet nuclei. A systematic review of the use of SFMs to reduce influenza transmission concluded that there are few data to endorse the wearing of a mask to prevent the wearer from becoming infected; there was in fact more evidence to suggest the use of a mask by an infected person can reduce transmission to others.⁵⁷ Studies comparing the effectiveness of SFMs and respirators are inconclusive to date and cannot easily be extrapolated to draw conclusions about modes of transmission.^{58–60}

The non-pharmaceutical intervention studies performed to reduce influenza transmission are summarised in Table 1. A problem with using interventions to assess modes of transmission is that blocking one route still allows transmission to take place down other alternative (unblocked/open) routes. For example, if contact transmission is blocked by HH, transmission could still occur via droplets and aerosols making the interpretation of any risk reduction complex.

Human influenza challenge studies

Humans can be experimentally infected with influenza following the instillation of drops intranasally⁶⁴ or by breathing aerosols.^{65,66} In the study by Alford, 23 volunteers inhaled 10L of an H2N2 aerosol delivered via a facemask. The dose of virus delivered ranged between 1 and 126 TCID₅₀. Four volunteers developed clinical illness; virus was isolated from these and one other volunteer, whilst seroconversion was seen in seven including all those who exhibited illness. Noting limitations of the study design and making an

assumption that only 60% of the aerosol load inhaled will reach the lower respiratory tract the study reports that half of the volunteers with very low pre-existing antibody titres were infected with 0.3–6 TCID₅₀. This is substantially lower than the infectious dose required for intranasal inoculation (100–1000 TCID₅₀)^{67–69} and has led some to conclude that the LRT is the preferred site of infection and by implication (as only aerosols can reach it) that the aerosol route of transmission is important.^{2,70} In addition, it has been suggested that natural infection may more closely resemble aerosol than intranasally initiated infection.⁷⁰

Animal models of transmission

The droplet and aerosol routes dominate in transmission experiments with animals (Table 2). Unfortunately, it is not possible to discriminate between them in most models^{71–74} although it has been argued that the experimental methods described favour the operation of aerosol over droplet transmission.⁷⁵ Aerosol inoculation of ferrets has been found to simulate natural infection more closely than intranasal inoculation, and viable virus has been detected in exhaled aerosols.⁷⁶ The contact transmission route has appeared less significant in animal studies; however, interpretation of this in the context of human to human transmission is problematic because of the markedly different social and physical behaviours of humans compared with small mammals. There seems little doubt that some environmental factors such as temperature and humidity can affect transmission between animals.^{77–79}

Through the use of animal models, a better understanding of the viral determinants of transmission is developing, although the variety and interplay of traits is complex; some seem to hinder transmission whilst others permit it through different routes. It is likely that viral properties (e.g. fitness for replication, receptor preferences) help determine infectiousness and modes of spread.^{80,81} However, the extent to which all findings can be generalised to human transmission is uncertain and scientifically challengeable.

Transmission modelling

A number of modelling scenarios have been constructed that combine defined physical dynamics with biologic processes to estimate outcomes (Table 3). Whilst most support the concept that all transmission routes can be important given the right circumstances, there appears to be divergence between those who conclude that droplet transmission is significant⁸³ and those who conclude it is less significant.^{27,84,85} Despite droplet particles having high infectivity potential, it is likely that their inability to reach target cells, and data which reveal that the infectious dose in the URT is higher than the LRT² compromise this. Some models suggest a significant role for contact transmission^{84–86} although

Table 1. Non-pharmaceutical intervention studies

Study (year)	Study design	Study aim (<i>n</i> = subjects analysed)	Study setting/ randomisation unit	Study arms	Main findings
Talaat (2010) ⁵⁵	Cluster randomised controlled trial	Primary prevention (<i>n</i> = 44 451)	Schools	HH/Control	Significant reductions in ILI absenteeism and laboratory-confirmed influenza (A+B)
Stebbins (2011) ⁵⁶	Cluster randomised controlled trial	Primary prevention (<i>n</i> = 3360)	Schools	Hand + Respiratory hygiene/Control	Significant reductions in absenteeism and laboratory-confirmed influenza A
Aiello (2008) ⁵⁴	Cluster randomised controlled trial	Primary prevention (<i>n</i> = 1297)	University residences	SFM/HH + SFM/Control	No difference in cumulative incidence of ILI. During study weeks 4–6, weekly ILI incidence reduced in SFM + HH versus control
Cowling (2009) ⁶¹	Cluster randomised controlled trial	Secondary prevention (<i>n</i> = 794)	Households (inc. index case)	HH/SFM + HH/Control	No difference in laboratory-confirmed secondary attack rates between study arms. Some effects seen if interventions (HH + SFM) implemented within 36 hours
MacIntyre (2009) ⁵⁹	Cluster randomised controlled trial	Secondary prevention (<i>n</i> = 286)	Households	SFM/respirator/Control	No difference in rate of ILI between arms. If compliant with mask use reductions in ILI seen with both masks
Larson (2010) ⁶²	Block randomised controlled trial	Primary and secondary prevention (<i>n</i> = 2788)	Households	HH/HH + SFM/Control	No difference in rates of clinical infection (upper respiratory infections and influenza). SFM use associated with reduced (SARs)
Simmerman (2011) ⁶³	Randomised controlled trial	Secondary prevention (<i>n</i> = 887)	Households	HH/SFM + HH/Control	No difference in laboratory-confirmed SAR between study arms.
Loeb (2009) ⁵⁸	Randomised controlled trial	Comparative non-inferiority (<i>n</i> = 446)	Healthcare workers (in hospitals)	SFM/Respirator	SFMs were non-inferior to respirators in relation to rates of laboratory-confirmed influenza
MacIntyre (2011) ⁶⁰	Cluster randomised controlled trial	Comparative efficacy (<i>n</i> = 1441)	Hospitals (healthcare workers)	SFM/Fit tested Respirator/Non-fit tested respirator	Respirators were associated with an approximate halving of risk for all infection outcomes compared with SFMs, but after adjustment for clustering, the only significant finding was that non-fit-tested respirators were more protective against clinical respiratory infection

HH, hand hygiene; ILI, influenza-like illness; LRT, lower respiratory tract; SFM, surgical face masks; URT, upper respiratory tract.

Table 2. Summary of animal studies

Author (year)	Study/Investigation	Main findings
Andrewes (1941) ⁷¹	Experimental infection in ferrets	Transmission occurred between ferrets housed in different cages and separated by distances that would arguably only permit aerosol spread
Schulman (1968) ⁷⁴	Experimental infection in mice	Transmission was demonstrated between mice housed in the same and separate cages, and the frequencies of transmitted infection were similar. One experiment allowed the ventilation in a cage housing donors and recipients to be altered; when ventilation was increased, infection rates decreased. These findings were interpreted as signifying that aerosol transmission was active
Lowen (2006) ⁷²	Transmission in guinea pigs	A human H3N2 virus was shown to replicate well in guinea pigs after intranasal inoculation and transmission from infected to recipient animals occurred when animals were housed together or in separate cages (side by side and separated by 91 cm)
Lowen (2007) ⁷⁸	Investigation of the effect of humidity and temperature on transmission in ferrets	Experiments on guinea pigs housed in an environmental chamber were conducted that only allowed for droplet or aerosol transmission. Low RH (20–30%) seemed to favour transmission whilst higher RH (80%) inhibited it. In another set of experiments, transmission occurred at low temperature (5°C) more frequently than higher temperatures (20 and 30°C)
Lowen (2008) ⁷⁹	Investigation of the effect of temperature on contact transmission in ferrets	Recipient guinea pigs were placed in cages that had housed infected ones with ambient temperatures of 20 and 30°C. Transmission was seen to occur equally at both temperatures; the authors suggest that whilst droplet and aerosol transmission is reduced by high temperatures, contact transmission is not (as virus is not released and therefore not exposed to the outside environment)
Mubareka (2009) ⁷³	Routes of transmission in guinea pigs	The relative contributions of droplet/aerosol and fomite (contact) transmission were studied. Infected and recipient animals were placed in separate cages >80 cm above each other and transmissions occurred. However, when recipient animals were placed in the cages of infected animals (infected animals were removed but fomites were not) less infection transmission was seen
Gustin (2011) ⁷⁶	Comparison of aerosol and intranasal inoculation of ferrets	An aerosol inoculation system was devised. Aerosol inoculation caused a more natural influenza infection. Onward transmission was dependent on the level and duration of virus shedding. Viable virus was detected from infected ferrets in exhaled aerosols
Roberts (2012) ⁸²	Investigation into pre-symptomatic transmission	Transmission occurred via both contact and respiratory droplet exposure before the earliest symptoms developed. Furthermore, transmission did not temporally correlate with respiratory symptoms, such as coughs and sneezes, but rather with the peak viral titre in the nose

Table 3. Modelling investigations

Author (year)	Study/Investigation	Main Findings
Atkinson (2008) ²⁷	Model to quantify role of aerosol, contact and droplet routes based on a household scenario	They conclude that aerosol transmission is far more dominant than contact transmission and that droplet transmission is difficult to analyse without reliable data on how often people are in close (<1 m) contact, their relative heights and the directions of emitted sneezes and coughs
Nicas (2008) ⁸⁶	Investigation of the hand-to-face contact route of transmission	The scenario was a caregiver attending a sick family member in a bedroom for 30 minutes. An infection risk due to hand contact of 0.011% was generated
Nicas (2009) ⁸⁴	Model to quantify routes of transmission based on a scenario of visiting a patient's room	Important variables were considered to be (i) infectious doses and (ii) viral titres. When the URT/LRT infectious dose ratio is 3200:1 contact, aerosol and droplet routes all contribute substantially to infection risk. With rising viral titres, contact and aerosol become more significant. When the ability of virus to reach target cells is taken into account, aerosol transmission assumes dominance
Wagner (2009) ⁹⁴	Risk assessment of aerosol transmission aboard an airplane	The authors find that proximity and duration of exposure to the source and passenger density are important factors. Up to 17 infections could be caused during a 17-hour flight
Spicknall (2010) ⁸⁵	Model to quantify routes of transmission	The indirect contact mode of transmission appeared dominant. However, the authors explain that this is not necessarily the case in all settings. Of 10 000 model runs, indirect contact was dominant in 3079, respiratory in 121 and droplet in 66. Furthermore, considerable overlap is also seen where modes appear co-dominant, this occurred in 1969 sets
Teunis (2011) ⁸³	Model to quantify routes of transmission	Infectious dose data from influenza challenge studies are used in the construct of a model that simulates infection from a patient in a poorly ventilated room. Infection droplets and aerosols are approximately equal

model outputs are highly dependent on estimates of infectious dose.

Asymptomatic and pre-symptomatic transmission

An important feature of infection in some individuals is that they shed virus but do not experience symptoms. This may happen early in the course of infection (pre-symptomatic) or exist throughout the course of an infection (asymptomatic). Such individuals may not seek treatment or self-isolate and therefore may be an important group. A recent ferret study has demonstrated that pre-symptomatic transmission does occur.⁸² Amongst humans, models have typically assumed that asymptomatic or subclinical infections make up 33–50% of all infections^{87,88} although empiric data obtained during the 2009 pandemic showed asymptomatic infection rates of 8–14%^{89–91} with a subclinical rate of 25%.⁹⁰ Lau *et al.*⁹⁰ estimated that 1–8% of infectiousness occurs prior to illness onset. However, the amount and duration of viral shedding from asymptomatic patients can be low^{90,92}, and it remains to be shown that asymptomatic humans effectively transmit influenza.⁹³

Conclusion

Contact

Contact transmission of influenza cannot be excluded; virus survival data show that it is biologically plausible. Its

importance, however, is questioned by field data, although the scarcity and uncertain quality of those data are themselves problematic issues. More data from infected patients in natural settings are needed.

Droplet

Droplet transmission is often assumed to be significant, probably because in epidemiologic investigations close proximity to the source patient is often noted to be necessary for transmission to occur; however, data generated from clinical studies to back this up are lacking. The issue is that close proximity spread does not adequately differentiate droplet transmission from other routes such as short-range aerosol transmission. Furthermore, despite the fact that the vast majority of virus released from an infected person during a cough or a sneeze is carried by droplets, with high infectious potential, droplets face two major challenges: (i) to reach their target cells and (ii) to satisfy the relatively high infectious dose needed to initiate infection in the URT (compared with the LRT).

Aerosol

Aerobiological studies reveal that inhalable infectious particles ($\leq 5 \mu\text{m}$) can be produced by patients and that virus can remain viable (and therefore potentially infectious) in these particles long enough to permit infection transmission. A role for aerosol transmission from some infected

individuals in the absence of known aerosol-generating procedures cannot be ruled out, and a lack of evidence of long-range influenza transmission is not adequate evidence of absence of aerosol transmission at shorter distances. Of all the routes, it is perhaps aerosol transmission that has received most interest over recent years; evidence (albeit mainly indirect) in support of the importance of its overall contribution is increasing, but is still not definitive.

At present, the existing evidence on influenza transmission supports a potential role for all routes of transmission. Their relative significance will depend on the set of circumstances acting at a given time. Dictating the process are factors related to the virus itself, the host and the environment. Transmission can likely occur through multiple routes during the same 'event'; it is a dynamic and opportunistic process.

Research needs relevant to policy and guidance

Further research is needed in the following areas to clarify policy and guidance issues^{95,96}:

1. studies that further determine the importance of proximity (range) on human–human transmission;
2. studies that improve current understanding about the heterogeneity of virus shedding between individuals and within the same individuals over time (and in relation to symptoms);
3. studies that clarify the aerosol-generating potential of individual procedures in healthcare settings;
4. studies that clarify the relative contribution of contact, droplet and aerosol transmission;
5. studies that clarify the importance of human–human transmission arising from contact with infected, asymptomatic and infected pre-symptomatic individuals; and
6. studies that determine the effectiveness/efficacy of different types of masks, HH and combinations of personal protective measures for reducing transmission of influenza.

Conclusion

At present, the existing evidence on influenza transmission supports a potential role for all routes of transmission. Their relative significance will depend on the set of circumstances acting at a given time. Dictating the process are factors related to the virus itself, the host and the environment. Transmission can likely occur through multiple routes during the same 'event'; it is a dynamic and opportunistic process.

Conflict of interest

BK has no conflict of interests to declare. JSN-V-T has received research funding from GlaxoSmithKline and F.

Hoffman-La Roche; and Astra-Zeneca within the last two years.

References

- 1 Brankston G, Gitterman L, Hirji Z, Lemieux C, Gardam M. Transmission of influenza A in human beings. *Lancet Infect Dis* 2007; 7:257–265.
- 2 Tellier R. Review of aerosol transmission of influenza A virus. *Emerg Infect Dis* 2006; 12:1657–1662.
- 3 Zambon MC. The pathogenesis of influenza in humans. *Rev Med Virol* 2001; 11:227–241.
- 4 Rogers GN, Paulson JC. Receptor determinants of human and animal influenza virus isolates: differences in receptor specificity of the H3 hemagglutinin based on species of origin. *Virology* 1983; 127:361–373.
- 5 Shinya K, Ebina M, Yamada S, Ono M, Kasai N, Kawaoka Y. Avian flu: influenza virus receptors in the human airway. *Nature* 2006; 440 (7083):435–436.
- 6 Weber TP, Stilianakis NI. Inactivation of influenza A viruses in the environment and modes of transmission: a critical review. *J Infect* 2008; 57:361–373.
- 7 Chan MC, Chan RW, Yu WC *et al.* Tropism and innate host responses of the 2009 pandemic H1N1 influenza virus in ex vivo and in vitro cultures of human conjunctiva and respiratory tract. *Am J Pathol* 2010; 176:1828–1840.
- 8 Olofsson S, Kumlin U, Dimock K, Arnberg N. Avian influenza and sialic acid receptors: more than meets the eye? *Lancet Infect Dis* 2005; 5:184–188.
- 9 Bischoff WE, Reid T, Russell GB, Peters TR. Transocular entry of seasonal influenza-attenuated virus aerosols and the efficacy of n95 respirators, surgical masks, and eye protection in humans. *J Infect Dis* 2011; 204:193–199.
- 10 Webster RG, Yakhno M, Hinshaw VS, Bean WJ, Murti KG. Intestinal influenza: replication and characterization of influenza viruses in ducks. *Virology* 1978; 84:268–278.
- 11 WHO. Review of latest available evidence on potential transmission of avian influenza (H5N1) through water and sewage and ways to reduce the risks to human health Geneva, 2007.
- 12 Bean B, Moore BM, Sterner B *et al.* Survival of influenza viruses on environmental surfaces. *J Infect Dis* 1982; 146:47–51.
- 13 Thomas Y, Vogel G, Wunderli W *et al.* Survival of influenza virus on banknotes. *Appl Environ Microbiol* 2008; 74:3002–3007.
- 14 McDevitt J, Rudnick S, First M, Spengler J. Role of absolute humidity in the inactivation of influenza viruses on stainless steel surfaces at elevated temperatures. *Appl Environ Microbiol* 2010; 76:3943–3947.
- 15 Grayson ML, Melvani S, Druce J *et al.* Efficacy of soap and water and alcohol-based hand-rub preparations against live H1N1 influenza virus on the hands of human volunteers. *Clin Infect Dis* 2009; 48:285–291.
- 16 Thomas YS, Peduzzi E, Eckert T, Koch D, Mathys P, Kaiser L. Survival of Influenza Virus on Human Fingers. Options for the control of influenza VII. Hong Kong: 2010.
- 17 Killingley B, Grotorex J, Cauchemez S *et al.* Virus shedding and environmental deposition of novel A (H1N1) pandemic influenza virus: interim findings. *Health Technol Assess* 2010; 14:237–354.
- 18 Simmerman JX, Suntarattiwong P, Levy J *et al.* Influenza virus contamination of common household surfaces during the 2009 influenza A (H1N1) pandemic in Bangkok, Thailand: implications for contact transmission. *Clin Infect Dis* 2010; 51:1053–1061.
- 19 Boone SA, Gerba CP. The occurrence of influenza A virus on household and day care center fomites. *J Infect* 2005; 51:103–109.

- 20 Bright KR, Boone SA, Gerba CP. Occurrence of bacteria and viruses on elementary classroom surfaces and the potential role of classroom hygiene in the spread of infectious diseases. *J Sch Nurs* 2010; 26:33–41.
- 21 Simmerman JM, Suntarattiwong P, Levy J *et al.* Influenza virus contamination of common household surfaces during the 2009 influenza A (H1N1) pandemic in Bangkok, Thailand: implications for contact transmission. *Clin Infect Dis* 2010; 51:1053–1061.
- 22 Suntarattiwong P, Levy J, Simmerman M *et al.* Influenza virus contamination of common household surfaces and its role in household transmission, Bangkok, Thailand. Oral Presentation A7020. European Scientific Working Group on Influenza. Malta: 2011.
- 23 Nicas M, Nazaroff WW, Hubbard A. Toward understanding the risk of secondary airborne infection: emission of respirable pathogens. *J Occup Environ Hyg* 2005; 2:143–154.
- 24 Chao CYH, Wan MP, Morawska L *et al.* Characterization of expiration air jets and droplet size distributions immediately at the mouth opening. *J Aerosol Sci* 2009; 40:122–133.
- 25 Gralton J, Tovey E, McLaws ML, Rawlinson WD. The role of particle size in aerosolised pathogen transmission: a review. *J Infect* 2010; 62:1–13.
- 26 Lidwell OM. The microbiology of air. Topley and Wilson's Principles of Bacteriology, Virology and Immunity. London: Hodder Arnold, 1990; 226–240.
- 27 Atkinson MP, Wein LM. Quantifying the routes of transmission for pandemic influenza. *Bull Math Biol* 2008; 70:820–867.
- 28 Frank AL, Taber LH, Wells CR *et al.* Patterns of shedding of myxoviruses and paramyxoviruses in children. *J Infect Dis* 1981; 144:433–441.
- 29 Lee N, Chan PK, Hui DS *et al.* Viral loads and duration of viral shedding in adult patients hospitalized with influenza. *J Infect Dis* 2009; 200:492–500.
- 30 Leekha S, Zitterkopf NL, Espy MJ *et al.* Duration of influenza A virus shedding in hospitalized patients and implications for infection control. *Infect Control Hosp Epidemiol* 2007; 28:1071–1076.
- 31 Sato M, Hosoya M, Kato K, Suzuki H. Viral shedding in children with influenza virus infections treated with neuraminidase inhibitors. *Pediatr Infect Dis J* 2005; 24:931–932.
- 32 Blachere FM, Lindsley WG, Pearce TA *et al.* Measurement of airborne influenza in a hospital emergency department. *Clin Infect Dis* 2009; 48:438–440.
- 33 Goyal SM, Anantharaman S, Ramakrishnan MA *et al.* Detection of viruses in used ventilation filters from two large public buildings. *Am J Infect Control* 2011; 39:e30–e38.
- 34 Lindsley WG, Blachere FM, Davis KA *et al.* Distribution of airborne influenza virus and respiratory syncytial virus in an urgent care medical clinic. *Clin Infect Dis* 2010; 50:693–698.
- 35 Yang W, Elankumaran S, Marr LC. Concentrations and size distributions of airborne influenza A viruses measured indoors at a health centre, a day-care centre and on aeroplanes. *J R Soc Interface* 2011; 8:1176–1184.
- 36 Milton DK, Fabian P, Angel M, Perez DR, McDevitt JJ. Influenza virus aerosols in human exhaled breath: particle size, culturability and effect of surgical masks. Swine origin H1N1: the first pandemic of the 21st century. Atlanta, USA, 2010.
- 37 Lindsley WG, Blachere FM, Thewlis RE *et al.* Measurements of airborne influenza virus in aerosol particles from human coughs. *PLoS ONE* 2010; 5:e15100.
- 38 Apisarnthanarak A, Mundy LM. Outbreak of influenza A (2009) H1N1 among Thai healthcare workers: is it time to integrate a vaccination program? *Infect Control Hosp Epidemiol* 2010; 31:854–856.
- 39 Awofeso N, Fennell M, Waliuzzaman Z *et al.* Influenza outbreak in a correctional facility. *Aust N Z J Public Health* 2001; 25:443–446.
- 40 Baker MG, Thornley CN, Mills C *et al.* Transmission of pandemic A/H1N1 2009 influenza on passenger aircraft: retrospective cohort study. *BMJ* 2010; 340:c2424.
- 41 Blumenfeld HL, Kilbourne ED, Louria DB, Rogers DE. Studies on influenza in the pandemic of 1957–1958. I. An epidemiologic, clinical and serologic investigation of an intrahospital epidemic, with a note on vaccination efficacy. *J Clin Invest* 1959; 38:199–212.
- 42 Cunney RJ, Bialachowski A, Thornley D, Smaill FM, Pennie RA. An outbreak of influenza A in a neonatal intensive care unit. *Infect Control Hosp Epidemiol* 2000; 21:449–454.
- 43 Han K, Zhu X, He F *et al.* Lack of airborne transmission during outbreak of pandemic (H1N1) 2009 among tour group members, China, June 2009. *Emerg Infect Dis* 2009; 15:1578–1581.
- 44 Klontz KC, Hynes NA, Gunn RA *et al.* An outbreak of influenza A/Taiwan/1/86 (H1N1) infections at a naval base and its association with airplane travel. *Am J Epidemiol* 1989; 129:341–348.
- 45 McLean RL. The effect of ultraviolet radiation upon the transmission of epidemic influenza in long term hospitals. *Am Rev Respir Dis* 1961; 83:36–38.
- 46 Morens DM, Rash VM. Lessons from a nursing home outbreak of influenza A. *Infect Control Hosp Epidemiol* 1995; 16:275–280.
- 47 Moser MR, Bender TR, Margolis HS *et al.* An outbreak of influenza aboard a commercial airliner. *Am J Epidemiol* 1979; 110:1–6.
- 48 Wong BC, Lee N, Li Y *et al.* Possible role of aerosol transmission in a hospital outbreak of influenza. *Clin Infect Dis* 2010; 51:1176–1183.
- 49 Cui F, Luo H, Zhou L *et al.* Transmission of pandemic influenza A (H1N1) virus in a train in China. *J Epidemiol* 2011; 21:271–277.
- 50 Piso RJ, Albrecht Y, Handschin P, Bassetti S. Low transmission rate of 2009 H1N1 Influenza during a long-distance bus trip. *Infection* 2011; 39:149–153.
- 51 Aledort JE, Lurie N, Wasserman J, Bozzette SA. Non-pharmaceutical public health interventions for pandemic influenza: an evaluation of the evidence base. *BMC Public Health* 2007; 7:208.
- 52 Jefferson T, Del Mar C, Dooley L *et al.* Physical interventions to interrupt or reduce the spread of respiratory viruses. *Cochrane Database Syst Rev* 2010; (1):CD006207.
- 53 Rabie T, Curtis V. Handwashing and risk of respiratory infections: a quantitative systematic review. *Trop Med Int Health* 2006; 11:258–267.
- 54 Aiello AE, Coulborn RM, Perez V, Larson EL. Effect of hand hygiene on infectious disease risk in the community setting: a meta-analysis. *Am J Public Health* 2008; 98:1372–1381.
- 55 Talaat MAS, Afifi S, Dueger E *et al.* Effects of hand hygiene campaigns on incidence of laboratory-confirmed influenza and absenteeism in schoolchildren, Cairo, Egypt. *Emerg Infect Dis* 2011; 17:619–625.
- 56 Stebbins S, Cummings DA, Stark JH *et al.* Reduction in the incidence of influenza A but not influenza B associated with use of hand sanitizer and cough hygiene in schools: a randomized controlled trial. *Pediatr Infect Dis J* 2011; 30:921–926.
- 57 Cowling BJ, Zhou Y, Ip DK, Leung GM, Aiello AE. Face masks to prevent transmission of influenza virus: a systematic review. *Epidemiol Infect* 2010; 138:449–456.
- 58 Loeb M, Dafeo N, Mahony J *et al.* Surgical mask vs N95 respirator for preventing influenza among health care workers: a randomized trial. *JAMA* 2009; 302:1865–1871.
- 59 MacIntyre CR, Cauchemez S, Dwyer DE *et al.* Face mask use and control of respiratory virus transmission in households. *Emerg Infect Dis* 2009; 15:233–241.
- 60 MacIntyre CR, Wang Q, Cauchemez S *et al.* A cluster randomized clinical trial comparing fit-tested and non-fit-tested N95 respirators to medical masks to prevent respiratory virus infection in health care workers. *Influenza Other Respi Viruses* 2011; 5:170–179.

- 61 Cowling BJ, Chan KH, Fang VJ *et al.* Facemasks and hand hygiene to prevent influenza transmission in households: a cluster randomized trial. *Ann Intern Med* 2009; 151:437–446.
- 62 Larson EL, Ferng YH, Wong-McLoughlin J *et al.* Impact of non-pharmaceutical interventions on URIs and influenza in crowded, urban households. *Public Health Rep* 2010; 125:178–191.
- 63 Simmerman JM, Sutarattiwong P, Levy J *et al.* Findings from a household randomized controlled trial of hand washing and face masks to reduce influenza transmission in Bangkok, Thailand. *Influenza Other Respi Viruses* 2011; 5:256–267.
- 64 Carrat F, Vergu E, Ferguson NM *et al.* Time lines of infection and disease in human influenza: a review of volunteer challenge studies. *Am J Epidemiol* 2008; 167:775–785.
- 65 Alford RH, Kasel JA, Gerone PJ, Knight V. Human influenza resulting from aerosol inhalation. *Proc Soc Exp Biol Med* 1966; 122:800–804.
- 66 Henle G, Stokes J, Maris EP. Experimental exposure of human subjects to viruses of influenza. *J Immunol* 1946; 52:145–165.
- 67 Hayden FG, Treanor JJ, Betts RF *et al.* Safety and efficacy of the neuraminidase inhibitor GG167 in experimental human influenza. *JAMA* 1996; 275:295–299.
- 68 Douglas R. *Influenza in Man*. New York: Academic Press, 1975.
- 69 Couch RB, Douglas RG Jr, Fedson DS, Kasel JA. Correlated studies of a recombinant influenza-virus vaccine. 3. Protection against experimental influenza in man. *J Infect Dis* 1971; 124:473–480.
- 70 Little JW, Douglas RG Jr, Hall WJ, Roth FK. Attenuated influenza produced by experimental intranasal inoculation. *J Med Virol* 1979; 3:177–188.
- 71 Andrewes C, Glover R. Spread of infection from the respiratory tract of the ferret. *Transmission of influenza A virus*. *Br J Exp Pathol* 1941; 22:7.
- 72 Lowen AC, Mubareka S, Tumpey TM, Garcia-Sastre A, Palese P. The guinea pig as a transmission model for human influenza viruses. *Proc Natl Acad Sci USA* 2006; 103:9988–9992.
- 73 Mubareka S, Lowen AC, Steel J *et al.* Transmission of influenza virus via aerosols and fomites in the guinea pig model. *J Infect Dis* 2009; 199:858–865.
- 74 Schulman JL. The use of an animal model to study transmission of influenza virus infection. *Am J Public Health Nations Health* 1968; 58:2092–2096.
- 75 Tellier R. Aerosol transmission of influenza A virus: a review of new studies. *J R Soc Interface* 2009; 6(Suppl 6):S783–S790.
- 76 Gustin KM, Belser JA, Wadford DA *et al.* Influenza virus aerosol exposure and analytical system for ferrets. *Proc Natl Acad Sci USA* 2011; 108:8432–8437.
- 77 Lowen AC, Palese P. Transmission of influenza virus in temperate zones is predominantly by aerosol, in the tropics by contact: a hypothesis. *PLoS Curr Influenza* 2009; 1:RRN1002.
- 78 Lowen AC, Mubareka S, Steel J, Palese P. Influenza virus transmission is dependent on relative humidity and temperature. *PLoS Pathog* 2007; 3:1470–1476.
- 79 Lowen AC, Steel J, Mubareka S, Palese P. High temperature (30 degrees C) blocks aerosol but not contact transmission of influenza virus. *J Virol* 2008; 82:5650–5652.
- 80 Belser JA, Maines TR, Tumpey TM, Katz JM. Influenza A virus transmission: contributing factors and clinical implications. *Expert Rev Mol Med* 2010; 12:e39.
- 81 Nicholls JM, Chan RW, Russell RJ, Air GM, Peiris JS. Evolving complexities of influenza virus and its receptors. *Trends Microbiol* 2008; 16:149–157.
- 82 Roberts KL, Shelton H, Stilwell P, Barclay WS. Transmission of a 2009 H1N1 pandemic influenza virus occurs before fever is detected, in the ferret model. *PLoS ONE* 2012; 7:e43303.
- 83 Teunis PF, Brienens N, Kretzschmar ME. High infectivity and pathogenicity of influenza A virus via aerosol and droplet transmission. *Epidemics* 2011; 2:215–222.
- 84 Nicas M, Jones RM. Relative contributions of four exposure pathways to influenza infection risk. *Risk Anal* 2009; 29:1292–1303.
- 85 Spicknall IH, Koopman JS, Nicas M *et al.* Informing optimal environmental influenza interventions: how the host, agent, and environment alter dominant routes of transmission. *PLoS Comput Biol* 2010; 6:e1000969.
- 86 Nicas M, Best D. A study quantifying the hand-to-face contact rate and its potential application to predicting respiratory tract infection. *J Occup Environ Hyg* 2008; 5:347–352.
- 87 Ferguson NM, Cummings DA, Fraser C *et al.* Strategies for mitigating an influenza pandemic. *Nature* 2006; 442:448–452.
- 88 Longini IM Jr, Halloran ME, Nizam A, Yang Y. Containing pandemic influenza with antiviral agents. *Am J Epidemiol* 2004; 159:623–633.
- 89 Sues T, Buchholz U, Dupke S *et al.* Shedding and transmission of novel influenza virus A/H1N1 infection in households—Germany, 2009. *Am J Epidemiol* 2010; 171:1157–1164.
- 90 Lau LL, Cowling BJ, Fang VJ *et al.* Viral shedding and clinical illness in naturally acquired influenza virus infections. *J Infect Dis* 2010; 201:1509–1516.
- 91 Papenburg J, Baz M, Hamelin ME *et al.* Household transmission of the 2009 pandemic A/H1N1 influenza virus: elevated laboratory-confirmed secondary attack rates and evidence of asymptomatic infections. *Clin Infect Dis* 2010; 51:1033–1041.
- 92 Cowling BJ, Chan KH, Fang VJ *et al.* Comparative epidemiology of pandemic and seasonal influenza A in households. *N Engl J Med* 2010; 362:2175–2184.
- 93 Patrozou E, Mermel LA. Does influenza transmission occur from asymptomatic infection or prior to symptom onset? *Public Health Rep* 2009; 124:193–196.
- 94 Wagner BG, Coburn BJ, Blower S. Calculating the potential for within-flight transmission of influenza A (H1N1). *BMC Med* 2009; 7:81.
- 95 Jones RM. Critical review and uncertainty analysis of factors influencing influenza transmission. *Risk Anal* 2011; 31:1226–1242.
- 96 Vukotich CJ Jr, Coulborn RM, Aragon TJ *et al.* Findings, gaps, and future direction for research in nonpharmaceutical interventions for pandemic influenza. *Emerg Infect Dis* 2010; 16:e2.